## Electronmicroscopical Contrast by Palladium Chloride

J. M. Ferrer, A. Tato, and J. C. Stockert

Departamento de Morfología Microscópica, Facultad de Biología, Universidad de Valencia, and Departamento de Citología e Histología, Facultad de Ciencias, Universidad Autónoma de Madrid, Spain.

Z. Naturforsch. 41c, 483-484 (1986); received December 20, 1985

Electron Dense Stain, Palladium Chloride, Chromatin, Eosinophil Granule

Thin sections of glutaraldehyde-fixed, epoxy resin-embedded bone marrow from rats were treated with 2% palladium chloride in 2% concentrated HCl. This procedure was found to induce high electron density in chromatin from all cell types and in cytoplasmic granules of neutrophils and eosinophils. In the latter, the crystalline body showed more contrast than the matrix.

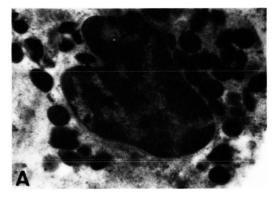
Contrast enhancement in electron microscopy (EM) usually involves "staining" with salts of heavy metals, the most frequently employed elements being U and Pb [1–3]. The use of other heavy metals as general or selective electron dense stains is also known, e.g. Fe, Cr, Co, Ru, In, Bi, Tl, Hg, Au, Ag, Os, Mo, W [2, 4–10]. However, elements from the platinum group (Pt, Pd) have scarcely been explored as contrasting agents for EM. Platinum was used as PtBr<sub>4</sub> [11], cis-dichloro-diammine-Pt(II) [12] and Pt-pyrimidine complexes [13], but no references were found in the case of palladium. We report here preliminary results on the use of palladium chloride as an electron stain.

Bone marrow from normal rats were fixed in 4% glutaraldehyde in 1/15 M Sörensen's buffer at pH 7.2 for 2 h and then rinsed in the same buffer for 2-3 h. Samples were dehydrated in acetone and embedded in Epon (Epikote 812) as usual. Thin sections were obtained with a Jum-7 (Jeol) ultramicrotome and transfered to gold grids without formvar. A 2% solution of palladium chloride (PdCl<sub>2</sub>, Merck) in 2% concentrated hydrochloric acid was prepared in distilled water. This solution, which corresponds to chloropalladious acid (H2PdCl4) in a small excess of HCl [14], was directly used for contrasting of sections. Treatments were made either at room temperature or at 35-40 °C for 2-3 h, after which sections were rinsed in distilled water and observed in a Jeol 100-S electron microscope operating at 60 KV.

Reprint requests to Prof. Dr. Juan C. Stockert, Departamento de Citología e Histología, Facultad de Ciencias, C-XV, Universidad Autónoma de Madrid, Canto Blanco, Madrid-28049, Spain.

Verlag der Zeitschrift für Naturforschung, D-7400 Tübingen  $0341-0382/86/0400-0483 \quad \$ \ 01.30/0$ 

Chromatin, specific granules of eosinophils and small round granules in neutrophils showed the most intense electron opacity after PdCl<sub>2</sub> treatment (Fig. 1). High density was observed in chromatin



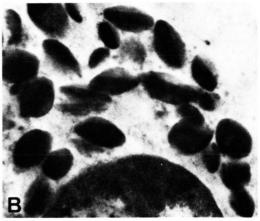


Fig. 1, A and B. Electron micrographs of eosinophil leucocytes showing the contrasting reaction of chromatin and cytoplasmic granules induced by  $PdCl_2$  treatment at 35 °C for 2 h. A:  $\times 12300$ ; B:  $\times 24200$ .



Dieses Werk wurde im Jahr 2013 vom Verlag Zeitschrift für Naturforschung in Zusammenarbeit mit der Max-Planck-Gesellschaft zur Förderung der Wissenschaften e.V. digitalisiert und unter folgender Lizenz veröffentlicht: Creative Commons Namensnennung-Keine Bearbeitung 3.0 Deutschland

This work has been digitalized and published in 2013 by Verlag Zeitschrift für Naturforschung in cooperation with the Max Planck Society for the Advancement of Science under a Creative Commons Attribution-NoDerivs 3.0 Germany License.

masses from all cell types present in bone marrow. The crystalline inclusion in eosinophil granules appeared more contrasted than the matrix, and a highly opaque boundary was often observed between them. Control sections (lacking PdCl<sub>2</sub> treatment) did not reveal electron density in these structures, and only diffuse outlines of cellular components were observed. Slight differences in the contrast degree were found in dependence on the temperature and duration of PdCl<sub>2</sub> treatment, the most intense contrast appearing after 3 h at 40 °C.

At present, it is difficult to explain the precise mechanism of this reaction. The reduction of Pd from oxidation state 2+ to zero by olefinic double bonds from lipids [14] does not seem to occur as no granular deposits of the metal were observed in EM. Pd(II) easily coordinate to nitrogen, oxygen and sulphur ligands [15], and chelate complexes of Pd with amino acids, their derivatives, and peptides [16–18], as well as pyrimidine and purine bases are known

- [1] C. R. Zobel and M. Beer, Int. Rev. Cytol. **18**, 363 (1965)
- [2] M. A. Hayat, Positive staining for electron microscopy. Van Nostrand Reinhold Co., New York 1975.
- [3] J. Derksen and H. Meekes, Micron **15**, 55 (1984).
- [4] J. M. Barret, P. M. Heidger, and S. W. Kennedy, J. Histochem. Cytochem. 23, 780 (1975).
- [5] A. Gautier, Int. Rev. Cytol. 44, 113 (1976).
- [6] J. C. Stockert, Histochemistry **53**, 43 (1977).
- [7] J. C. Stockert, Biol. Cell. 29, 211 (1977).
- [8] G. Moyne, Progr. Histochem. Cytochem. 13, 1 (1980).
- [9] J. C. Stockert and R. Paniagua, Z. Naturforsch. 35c, 348 (1980).
- [10] J. M. Ferrer, A. Tato, and J. C. Stockert, J. Microscopy 134, 221 (1984).
- [11] H. Finck, J. Biophys. Biochem. Cytol. 4, 291 (1958).

[19]. It is also reasonable to expect that the chloropalladite anion  $(PdCl_4)^{2-}$  would bind to positively charged groups in tissues [14]. Reactions of  $PdCl_2$  with nitrogen ligands from DNA, histones, and other basic proteins could account for the electron opacity of chromatin and cytoplasmic granules. In addition, these structures are compact and poorly infiltrated by epoxy resins, which allows an easier penetration of aqueous reagents [20]. Although further investigations are required to determine more precisely the contrasting mechanism and selectivity by  $PdCl_2$ , these results show that Pd (atomic weight 106.4) can be applied as an interesting metal to enhance contrast in EM.

## Acknowledgements

This work was supported by a grant (2320-83) from the Comisión Asesora de Investigación Científica y Técnica, Spain.

- [12] E. Heinen, Histochemistry **51**, 267 (1977).
- [13] S. K. Aggarwal, J. Histochem. Cytochem. 24, 984 (1976).
- [14] J. A. Kiernan, J. Histochem. Cytochem. **25**, 200 (1977).
- [15] F. R. Hartley, The chemistry of platinum and palladium. Applied Science Publishers, London 1973.
- [16] U. Taubald, U. Nagel, and W. Beck, Chem. Ber. 117, 1003 (1984).
- [17] A. Ryabov, V. A. Polyakov, and A. K. Yatsimirsky, Inorg. Chim. Acta 91, 59 (1984).
- [18] S. H. Kim and R. B. Martin, J. Amer. Chem. Soc. 106, 1707 (1984).
- [19] S. H. Kim and R. B. Martin, Inorg. Chim. Acta **91**, 11 (1984).
- [20] J. C. Stockert, Micr. Electr. Biol. Cel. 8, 159 (1985).